

PII: S0960-894X(96)00580-X

SYNTHESIS AND BIOLOGICAL EVALUATION OF 5,6-DIARYLIMIDAZO[2.1-b]THIAZOLE AS SELECTIVE COX-2 INHIBITORS

Michel Thérien,* Christine Brideau, Chi Chung Chan, Wanda A. Cromlish, Jacques Yves Gauthier, Robert Gordon, Gillian Greig, Stacia Kargman, Cheuk Kun Lau, Yves Leblanc, Chun-Sing Li, Gary P. O'Neill, Denis Riendeau, Patrick Roy, Zhaoyin Wang, Lijing Xu, and Petpiboon Prasit

Merck Frosst Centre for Therapeutic Research P.O. Box 1005, Pointe Claire - Dorval, Québec, H9R 4P8, Canada.

Abstract: A series of 5,6-diarylimidazo[2.1-b]thiazole compounds were prepared and their inhibitory potencies against COX-2 and Cox-1 enzymes were measured. This led to the identification of L-766,112 as a potent, orally active and selective inhibitor of the COX-2 enzyme. Copyright © 1996 Elsevier Science Ltd

Prostaglandins (PGs) are well known to be mediators of inflammation, pain and swelling.¹ They are produced by the action of the cyclooxygenase (COX) enzyme on arachidonic acid. Todays nonsteroidal antiinflammatory drugs (NSAIDs) act through the inhibition of the COX reaction. However, the fact that some PGs are cytoprotective in the gastrointestinal tract severely limits the usefulness of these NSAIDs because of side effects such as ulceration, perforation and hemorrhage.² The recent discovery of an inducible isoform of cyclooxygenase (COX-2)³ that is associated primarily with inflammation has led to the hypothesis that the antiinflammatory effect of NSAIDs is due to the inhibition of COX-2 and that NSAIDs induced GI toxicity may be caused by the inhibition of the cytoprotective COX-1 enzyme and that has obviously produced significant interest. Compounds that selectively inhibit the COX-2 enzyme have been shown to possess antiinflammatory properties and reduced ulcerogenicity^{4a,b} in animal models and would have tremendous therapeutic potential if these properties translated in humans.

Several research groups are actively pursuing the development of COX-2 selective inhibitor.⁴ A number of classes of selective COX-2 inhibitors have emerged such as the tricyclic class best represented by the cyclopentene class,^{5a} Dup 697,^{5b} the sulfonamide class with NS-398,^{5c} Flosulide^{5d} and L-745,337,^{5e} and the indole class with L-761,066.^{5f} Recently, Gauthier^{4a} and Leblanc,^{6a} from our laboratory have reported the study of the substitution and replacement of the thiophene ring of Dup 697. As a further diversification of the heterocyclic template, we report the identification of L-766,112 (**2a**) as a potent, orally active and selective COX-2 inhibitor, where the thiophene ring of Dup 697 is replaced by an imidazo[2.1-b]thiazole bicyclic system.

$$Br$$
 SO_2Me H N SO_2Me H N SO_2Me F MeO $NS-398$ $X = O$ Flosulide $X = S$ $L-761,066$

Synthesis

The 5,6-diarylimidazo[2.1-b]thiazoles were prepared by the condensation of α-bromodiarylethanones 16 with appropriately substituted 2-aminothiazoles (all commercially available) in refluxing ethanol or isopentanol⁷ (Scheme 1). The carboxylic acid **2p** was obtained by the hydrolysis of ester **2o** (Scheme 2), which was prepared according to Scheme 1. Chlorinated analog **2q** was prepared by treatment of compound **2a** with sulfuryl chloride followed by DBU. Upon treatment of **2q** with sulfuryl chloride the trichloro-compound **2r** was obtained (Scheme 3).

Scheme 1

R¹ = H, SMe, SO₂Me, F
R² = H, SMe, SO₂Me
R³
$$R^4$$
 = H, Me, COOMe, COOH,Ph

Scheme 2

Scheme 3

Table I: Inhibition of Cox-2 and Cox-1 enzyme by 5,6-diarylimidazo[2.1-b]thiazole.

Compd	R ¹	R ²	R ³	R ⁴	R ⁵	COX-2 (IC ₅₀ μM)	COX-1 (IC ₅₀ μM)
2a	Н	Н	4-MeSO ₂	Н	Н	0.016	>50
2b	Н	Н	Н	4-MeS	Н	5.0	0.078
2c	Н	Н	Н	4-MeSO ₂	Н	3.21	>50
2d	Н	Н	4-MeS	Н	Н	0.42	0.32
2 e	Me	Н	Н	$4-MeSO_2$	Н	0.14	>50
2f	Н	Н	4-MeSO ₂	F	Н	0.014	>50
2g	Н	Н	4-MeSO ₂	F	F	0.012	>50
2h	Me	Н	4-MeSO ₂	Н	Н	0.012	>50
2i	Н	Me	4-MeSO ₂	Н	Н	3.0	>50
2j	Me	Me	4-MeSO ₂	Н	Н	5.0	>50
2k	Me	Me	Н	4-MeSO ₂	Н	>5.0	>50
21	Н	Me	Н	4-MeSO ₂	Н	1.0	>50
2m	-CH=CH-CH=CH-		4-MeSO ₂	Н	Н	>5.0	>50
2n	-СН=СН-СН=СН-		Н	4-MeSO ₂	Н	>5.0	>50
20	Н	CH ₂ COOEt	4-MeSO ₂	Н	Н	0.9	>50
2p	Н	CH ₂ COOH	4-MeSO ₂	Н	Н	>5	>50
2 q	C1	Н	4-MeSO ₂	Cl	Н	0.016	>50
2r	Cl	Cl	4-MeSO ₂	Cl	Н	3.6	>50

IC₅₀ values for the inhibition of the production of PGE₂ in arachidonic acid stimulated chinese hamster ovary (CHO) cells transfected with human COX-1 or COX-2.8

Discussion

In our pursuit of a selective COX-2 inhibitor, we have studied extensively the replacement of the thiophene center ring of Dup 697 with various 5- and 6-membered heterocycles. Our attention was then turned towards 5,6- and 5,5-bicyclic systems. A number of different bicyclic systems were prepared and evaluated and the imidazo[2.1-b]thiazole turned out to be one of the most potent and selective series of COX-2 inhibitors. Compounds 2a-2r were tested in vitro for their ability to inhibit the arachidonic acid-dependent production of PGE2 mediated by the COX-1 and COX-2 enzymes in chinese hamster ovary cells⁸ as the primary screen and the results are reported in Table I. In general the SAR was quite tight. As with other tricyclic inhibitors^{6b} of the same class the methyl sulfone proved to be essential for good COX-2 activity and selectivity, demonstrated with compounds 2a vs. 2d and 2c vs. 2b. All compounds other than 2b and 2d were inactive against COX-1 under the conditions evaluated.

In general, compounds with the 4-methylsulfonylphenyl substituent at the 6-position were not very active against the COX-2 enzyme except for compound 2e. Compounds that were unsubstituted at the 2- and 3position, (2a, 2f, and 2g) were quite potent and selective (IC₅₀ = 0.016, 0.014 and 0.016 μ M vs. COX-2, respectively). Monosubstitution at the 2-position gave compounds 2e and 2h, which retained their potency (0.039 and 0.012 μM, respectively, against Cox-2). However, substitution at the 3-position resulted in a significant loss of potency (2i, 2j, 2k, and 2l have an IC₅₀ of 3.0, 5.0, >5.0, and 1.0 μM, respectively). Compounds 2m and 2n, with a fused phenyl ring at the 2,3-position, are also inactive, which is expected since substituents at the 3-position seem to be detrimental to the COX-2 inhibitory potency. Similarly, an acetate substituent at the 3-position (20 and 2p), caused a major decrease in potency against COX-2. Among the compounds described in Table I, four of these stand out as candidates for further evaluation, these are compounds 2a, 2h, 2f, and 2g. They possess COX-2 activity similar to those of Dup 697 and Indomethacin. However, their selectivity is superior (Dup-697 is 0.002 and 0.059 µM against COX-2 and, COX-1, and Indomethacin is 0.026 and 0.018 against COX-2 and, COX-1, respectively). Measuring their ability to inhibit the COX-2 enzyme in human whole blood identifies compounds 2a and 2f (IC₅₀ = 0.9 and 0.4 μ M, respectively) as clearly superior to compounds 2h and 2g (IC₅₀ = 17.5 and 2.2 μ M, respectively), this difference in activity might be due to protein binding in the whole blood. Also, in the COX-1 human whole blood assay these compounds, 2a and 2f, proved to be more selective with $IC_{50} > 30 \mu M$ for 2a and $IC_{50} = 25$ μM for 2f. With an ED₅₀ = 2mg/kg vs. 5 mg/kg in the carrageenin induced rat paw edema assay, compound 2a was shown to be slightly superior to compound 2f and also superior to Dup-697 (57% inhibition at 10 mg/kg)10 in this model. In a GI permeability assay, when rats were dosed with compound 2a at 100 mg/kg bid for 10 days there was no increase in the level of 51Cr excreted in the feces11, indicative of a high level of GI tolerance. Unfortunately, compound L-766,112 was found to be inactive in an adjuvant arthritis model, which might be

due to the low bioavailability, where C_{max} is 1 μ M at 1 and 2 h, in rats dosed P.O. at 20 mg/kg. Furthermore, metabolism studies of L-766,112¹² (2a) have shown that the oxidation of the sulfur atom at the 1 position of the heterocyclic system gave a reactive metabolite that may result in potential problems. In order to address the oxidative metabolism problems, compounds (2q and 2r) with electron withdrawing substituents on the heterocyclic rings were prepared. Indeed, introduction of electron withdrawing chloride substituents did significantly reduce metabolism since after incubation with rat hepatocytes compounds 2q and 2r were nearly quantitatively recovered, whereas 2a is extensively metabolized. Unfortunately, the in vitro potency of compound 2q did not translate in vivo and compound 2r was not found to be sufficiently active.

In conclusion, we have identified a new series of selective COX-2 inhibitors that are active in vitro and in vivo. We have also identified L-766,112 as a very potent and selective COX-2 inhibitor. This potency was partially translated in vivo as shown by its activity in the rat paw edema assay. However, the poor metabolic profile of L-766,112 prompted us to investigate other tetracyclic systems. Further studies on the metabolism of L766,112 and on improving the in vivo potency are presented in the following papers.

References

- (a) Vane, J. R. Nature (London) 1971, 231, 232. (b) Smith, J. B.; Willis, A. L. Nature (London) 1971, 231, 235.
- 2. Allison, M. C.; Howatson, A. G.; Torrance, C. J.; Lee, F. D.; Russell, R. I. New Engl. J. Med. 1992, 327, 749
- (a) Mitchell, J. A.; Akarasereenont, P.; Thienermann, C.; Flower, R. J.; Vane, J. R. Proc. Natl. Acad. Sci. U.S.A. 1993, 90, 11693. (b) Meade, E. A.; Smith, W. L.; DeWitt, D. L. J. Biol. Chem. 1993, 268, 6610. (c) O'Neill, G. P.; Mancini, J. A.; Kargman, S.; Yergey, J.; Kwan, M. Y.; Falgueyret, J. P.; Abramovitz, M.; Kennedy, B. P.; Ouellet, M.; Cromlish, W.; Culp, S.; Evans, J. F.; Ford-Hutchinson, A. W.; Vickers, P. J. Mol. Pharmacol. 1994, 45, 245.
- (a) Gauthier, J. Y.; Leblanc, Y.; Black, W. C.; Chan, C. C.; Cromlish, W. A.; Gordon, R.; Kennedy, B. P.; Lau, C. K.; Leger, S.; Wang, Z.; Ethier, D.; Guay, J.; Mancini, J.; Riendeau, D.; Tagari, P.; Vickers, P.; Wong, E.; Xu, L.; Prasit, P. Bioorg. Med. Chem. Lett. 1996, 6, 87. (b) Black, W. C.; Bayly, C.; Belley, M.; Chan, C. C.; Charleson, S.; Denis, D.; Gauthier, J. Y.; Gordon, R.; Guay, D.; Kargman, S.; Leblanc, Y.; Mancini, J.; Ouellet, M.; Percival, D.; Roy, P.; Skorey, K.; Tagari, P.; Vickers, P.; Wong, E.; Xu, L.; Prasit, P. Bioorg. Med. Chem. Lett. 1996, 6, 725. (c) For a review see: Reitz, D. B.; Seibert, K. Annu. Rep. Med. Chem. 1995, 30, 179.

- (a) Reitz, D. B.; Li. J. J.; Norton, M. B.; Reinhard, E. J.; Collins, J. T.; Anderson, G. D.; Gregory, S. A.; Koboldt, C. M.; Perkins, W. E.; Seibert, K.; Isakson, P. C. J. Med. Chem. 1994, 37, 3878. (b) Gans, K. R.; Galbraith, W.; Roman, R. J.; Haber, S. B.; Kerr, J. S.; Schmidt, W. K.; Smith, C.; Hewes, W. E.; Ackerman, N. R. J. Pharmacol. Exp. Ther. 1990 254, 180. (c) Futaki, N.; Takahashi, S.; Yokoyama, M.; Arai, I; Higuchi, S.; Otomo, S. Prostaglandins, 1994, 47, 55. (d) Wiesenberg-Böttcher, I.; Schweizer, A.; Green, J. R.; Seltenmeyer, Y.; Müller, K. Agents Actions 1989, 26, 240. (e) Li. C. S.; Black, W. C.; Chan, C. C.; Ford-Hutchinson, A. W.; Gauthier, J. Y.; Gordon, R.; Guay, D.; Kargman, S.; Lau, C. K.; Mancini, J.; Ouimet, N.; Roy, P.; Vickers, P.; Wong, E.; Young, R. N.; Zamboni, R.; Prasit, P. J. Med. Chem. 1995, 38, 4897. (f) Leblanc, Y.; Black, W. C.; Chan, C. C.; Charleson, S.; Delorme, D.; Denis, D.; Gauthier, J. Y.; Grimm, E. L.; Gordon, R.; Guay, D.; Hamel, P.; Kargman, S.; Lau, C. K.; Mancini, J.; Ouellet, M.; Percival, D.; Roy, P.; Skorey, K.; Tagari, P.; Vickers, P.; Wong, E.; Xu, L.; Prasit, P. Bioorg. Med. Chem. Lett. 1996, 6, 731.
- (a) Leblanc, Y.; Gauthier, J. Y.; Ethier, D.; Guay, J.; Mancini, J.; Riendeau, D.; Tagari, P.; Vickers, P.;
 Wong, E.; Prasit, P. Bioorg. Med. Chem. Lett. 1995, 5, 2123. (b) Haber, S. B. U.S. Patent 4 820 827 (1989)
- 7. Werbel, L. M.; Zamora, M. L. J. Het. Chem. 1965, 2, 287.
- 8. Kargman, S.; Wong, E.; Greig, G.; Falgueyret, J. P.; Cromlish, W.; Ethier, D.; Yergey, J.; Riendeau, D.; Evans, J. F.; Kennedy, B.; Tagari, P.; Francis, D.; O'Neill, G. P. *Biochem. Pharmacol.* **1996**, *52*, 1113.
- 9. Brideau, C.; Kargman, S.; Liu, S.; Dallob, A. L.; Ehrich, E. W.; Rodger, I. W.; Chan, C. C. *Inflamm. Res.* 1996, 45, 68.
- Bertenshaw, S. R.; Talley, J. J.; Rogier, D. J.; Graneto, M. J.; Rogers, R. S.; Kramer, S. W.; Penning, T. D.;
 Koboldt, C. M. Veenhuizen, A. W.; Zhang, Y.; Perkins, W. E. Bioorg. Med. Chem. Lett. 1995, 5, 2919.
- Chan, C. C.; Boyce, S.; Brideau, C.; Ford-Hutchinson, A. W.; Gordon, R.; Guay, D.; Hill, R. G.; Li, C. S.;
 Mancini, J.; Panneton, M.; Prasit, P.; Rasori, R.; Riendeau, D.; Roy, P.; Tagari, P.; Vickers, P.; Wong, E.;
 Rodger, I. W. J. Pharmacol. Exp. Ther. 1995, 274, 1531.
- 12. See following paper.

(Received in USA 17 September 1996; accepted 25 November 1996)